



In vitro* antifungal effect of phenylboronic and boric acid on *Alternaria alternata

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The ascomycete fungus *Alternaria alternata* causes early blight, one of economically the most important tomato diseases. Due to frequent use of fungicides, *A. alternata* has developed resistance with negative economic and environmental consequences. Research of new ways to control fungal pathogens has turned its eye to environmentally friendly chemicals with low toxicity such as boronic acids. The aim of our study was therefore to test the antifungal effects of phenylboronic and boric acid *in vitro* on *A. alternata*. We isolated the pathogen from a symptomatic tomato plant and determined the minimum inhibitory concentration of phenylboronic and boric acid on *A. alternata* mycelial growth using the poisoned food technique. The antifungal effect was tested on a wide range of phenylboronic and boric acid concentrations (from 0.04 % to 0.3 %) applied separately to agar with mycelial disc of the pathogen. After five days of incubation, phenylboronic acid at low concentration (0.05 %) completely inhibited mycelial growth. Boric acid, in turn, did not significantly slow down mycelial growth but did reduce sporulation and confirmed its fungistatic effect. Our findings point to the potential use of phenylboronic acid to control phytopathogenic fungi. This is, to our knowledge, the first report on its antifungal effect on an agriculturally important pathogen *in vitro*. Moreover, since *A. alternata* is also a human pathogen, these results may have clinical ramifications.

KEY WORDS: boronic acids; early blight; minimal inhibitory concentration; mycelial growth; sporulation; tomato

Some species of phytopathogenic fungi have developed strong resistance to agrochemicals used for their control, which raises concern about the negative economic and environmental consequences in global food production (1, 2). The ascomycete fungus *Alternaria alternata* (Fr.) Keissler is of particular concern, as it causes early blight in tomato and incurs basal stem lesions on seedlings, stem lesions on adult plants, and fruit rot (3, 4), resulting in significant losses in crop yields (up to 79 %) and subpar nutritional quality. Furthermore, *A. alternata* and other *Alternaria* species produce mycotoxins (5) and pose a serious health threat to humans and livestock (6–10).

In vitro tests have revealed that several isolates of *A. alternata* are resistant to pyraclostrobin, boscalid, strobilurine (11), and azoxystrobin (12). In addition, consumers are increasingly concerned about pesticide residues in food, which is why fungicides will not be the future first choice for controlling fungal diseases (13). Recent years have seen a growing interest for environmentally friendly alternatives, and boronic acids have caught the eye of the scientific community, as they can inhibit a wide range of fungi (14) and are not toxic for the environment (15). This particularly concerns boric acid (BA), a common disinfectant (16), and its phenyl derivative, phenylboronic acid (PBA), a commercially available chemical with

antimicrobial (17, 18), antitumor (19), antibacterial (20, 21), and antifungal properties (22, 23) confirmed against several species of human fungi (24, 25). PBA is not toxic to the environment (18, 26–28), while boron is in fact an essential micronutrient for plants (29).

However, no research has yet investigated PBA activity against pathogenic fungi that attack agriculturally interesting plants, and the aim of this study was to address this gap by testing antifungal effects of PBA and BA on *A. alternata*.

MATERIALS AND METHODS

***A. alternata* isolation**

Tomato (*Solanum lycopersicum* cv. Rutgers) leaves with early blight symptoms were collected in the field and the infected plant material was incubated on potato dextrose agar (PDA, Sigma-Aldrich, St. Louis, MO, USA) in a climate chamber at 25 °C for five days as detailed by Nagrale et al. (30) to stimulate fungal growth. The obtained pure culture of the isolated fungi was determined morphologically (30) and with polymerase chain reaction (PCR) (31).

Preparation of PBA and BA in concentration range

PBA (Sigma-Aldrich, CAS No. 98-80-6) and BA (Sigma-Aldrich, CAS No. 10043-35-3) were used in a wide range of concentrations (0.04 %, 0.05 %, 0.06 %, 0.07 %, 0.08 %, 0.09 %, 0.1%, 0.2 % and 0.3 %, which corresponds to 0.4 mg/mL to 3.0 mg/mL). Five hundred milligrams of PBA or BA were dissolved in 50 mL of sterile distilled water to give a 1 % PBA or BA stock solution. Based on the dilution factor, an appropriate volume of 1 % PBA or BA solution was pipetted into 50 mL of melted PDA nutrient medium, which was poured in three Petri dishes for three repetitions.

Determination of minimum inhibitory concentrations

We used the poisoned food technique (32) with slight modifications (33) to determine the minimum inhibitory concentration (MIC) of PBA and BA on mycelial growth of *A. alternata*. Melted PDA agar with a varying PBA or BA concentrations was poured onto three plates for each concentration. Mycelial discs of *A. alternata* with a diameter of 5 mm were cut with a circular cutter and placed in the centre of the solidified PDA plates. Mycelial discs were assessed under a stereomicroscope (SZ 4045, Olympus, Tokyo, Japan) at 250x magnification. Control Petri dishes did not contain PBA or BA. Petri dishes were incubated in a climate chamber at 25 °C for five days to allow *A. alternata* colonies to develop enough for us to quantify the antifungal effect of PBA or BA.

Grown fungal colonies of *A. alternata* in Petri dishes were photographed on the colony counter (Scan 100, Interscience, France) and the obtained photos processed with the *ImageJ* open-source software (US National Institutes of Health, Bethesda, Maryland, USA) (34) according to Guzmán et al. (35). The growth of *A. alternata* was quantified by measuring the surface area of the grown colony and calculating the mean of three repetitions.

Statistical analysis

Mean surface areas of fungal colonies treated with PBA or BA were compared with control using the one-way analysis of variance (ANOVA), followed by Tukey's test to identify significant differences ($P < 0.05$).

RESULTS AND DISCUSSION

Antifungal activity of PBA against *A. alternata*

The antifungal effect of PBA on *A. alternata* mycelial growth is shown in Table 1 and Figure 1. After five days of incubation at 25 °C, no pathogen growth was observed at concentrations ranging from 0.05 % to 0.3 %, whereas 0.04 % PBA reduced fungal growth by 98 % compared to control. Mycelial discs showed no hyphal growth. Fungal growth reduction was statistically significant at all tested PBA concentrations compared to control (Tukey's test, $P < 0.05$), which points to highly effective antifungal activity against *A. alternata*. These results support earlier *in vitro* findings reported by Liu et al. (23), in which the application of 0.3 % PBA completely inhibited the growth and development of basidiomycete fungi that cause decay of Japanese cedar wood. In fact, in our study *A. alternata* has shown much higher sensitivity, as its growth was completely inhibited at much lower PBA concentrations, starting with 0.05 %.

Antifungal activity of BA against *A. alternata*

Unlike PBA, BA did not completely inhibit the mycelial growth of *A. alternata* after five days of incubation (Table 2 and Figure 2). Highest inhibition was achieved with mid-range concentrations, and the non-linear relationship between BA concentrations and mycelial growth may point to experimental variation.

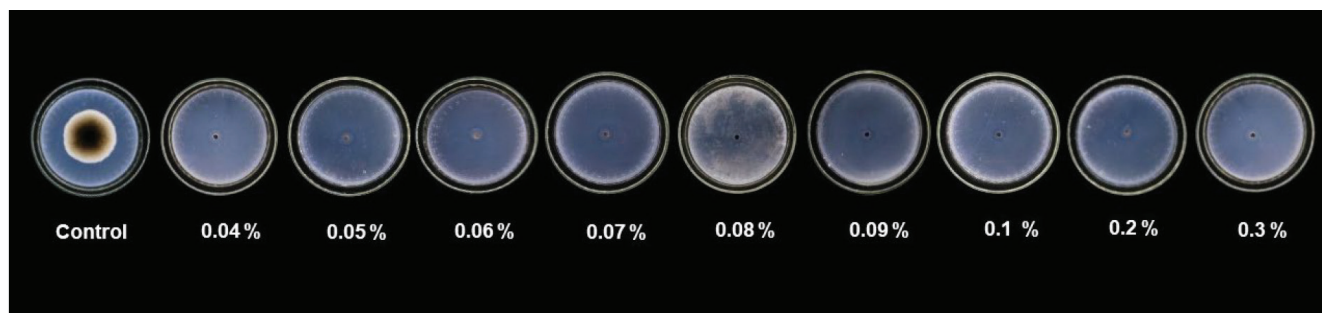


Figure 1 Effect of different doses of phenylboronic acid (volume concentration, %) on mycelial growth of pathogen *Alternaria alternata* on potato dextrose agar after 5 days of incubation at 25 °C

Table 1 Mycelial-growth area of *Alternaria alternata* after five days of incubation on potato dextrose with different phenylboronic acid volume concentrations (%) at 25 °C

PBA concentration (%)	0	0.04	0.05	0.06	0.07	0.08	0.09	0.1	0.2	0.3
Mean of colony area \pm SD	17.7 \pm 0.5 ^b	0.33 \pm 0.1 ^a	0 ^a	0 ^a	0 ^a	0 ^a	0 ^a	0 ^a	0 ^a	0 ^a

Values are presented as means \pm SD. Means with the same superscript letters across columns are not significantly different (One-way ANOVA; Tukey's test, $P < 0.05$)

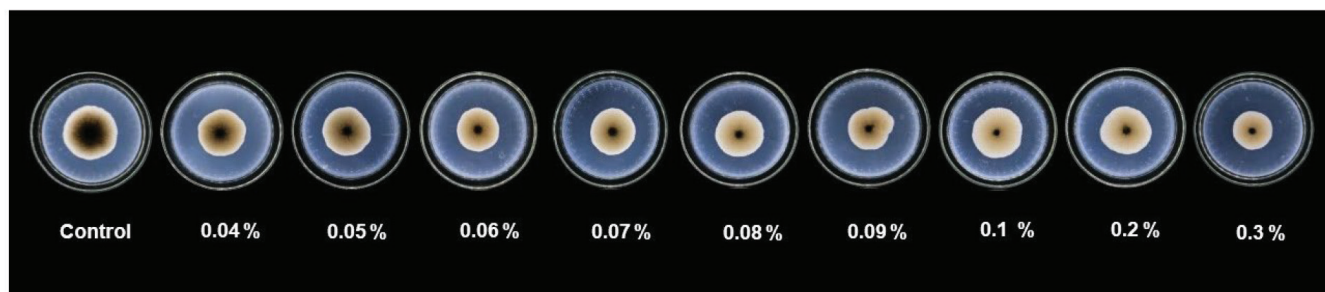


Figure 2 Effect of different doses of boric acid (volume concentration, %) on mycelial growth of pathogen *Alternaria alternata* on potato dextrose agar after 5 days of incubation at 25 °C

Table 2 Mycelial-growth area of *Alternaria alternata* after five days of incubation on potato dextrose agar with different boric acid concentrations at 25 °C

BA concentration (%)	0	0.04	0.05	0.06	0.07	0.08	0.09	0.1	0.2	0.3
Mean of colony area \pm SD	17.7 \pm 0.5 ^c	12.9 \pm 1.1 ^{ab}	11.2 \pm 0.7 ^{ab}	9.7 \pm 0.4 ^a	11.8 \pm 1 ^{abc}	12.2 \pm 0.5 ^{ab}	11.2 \pm 1.3 ^{ab}	14 \pm 0.1 ^{bc}	14.9 \pm 1 ^{abc}	16.2 \pm 0.6 ^{bc}
Inhibition (%)	0	27.1	36.7	45.2	33.3	31.1	36.7	20.9	15.8	8.5

Values are presented as means \pm SD. Means with the same superscript letters across columns are not significantly different (one-way ANOVA; Tukey's test, $P < 0.05$)

Morphological characteristics of the mycelia grown on media supplemented with BA differed from control. The mycelium was less coloured and less branched than the dark brown mycelium that grew on control substrate. This could be because BA at mid-range concentrations reduced sporulation without inhibiting significantly mycelial growth. Considering that *A. alternata* is a foliar fungus whose pathogenicity depends on sporulation and the way its conidia spread, this effect on sporulation is important for arresting or inhibiting pathogenesis.

Our findings are in line with one early report of *A. alternata* being tolerant to BA (36), whereas one study reported diminishing inhibitory effects on the germination of spores in *Alternaria* spp. from 44 % to 36 % over 48 h as BA concentrations rose from 0.2 % to 1 % (37). In contrast, another *in vitro* study (38) reported rising growth inhibition in *A. solani* with BA concentration over seven days of incubation, namely 90.16 % with 1 % and 95.22 % with 2 %

In an extension of this study, we investigated *in vivo* activity of BA and PBA against *A. alternata* infection of tomato plants in the same concentration ranges, which confirmed stronger prophylactic activity of PBA, in controlling early blight symptoms in test plants (28).

CONCLUSION

This study has established that PBA completely inhibits the growth of *A. alternata* at quite low doses, while BA inhibits sporulation. To our knowledge, this is the first report about *in vitro* antifungal activity of PBA against this agriculturally important pathogen and mycotoxin producer. Since *A. alternata* is also a human

pathogen, this study has potential pharmaceutical ramifications, especially as PBA is well tolerated by mammals (19, 27).

Another advantage of PBA is that it is environmentally friendly.

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Conflicts of interest

None to declare.

REFERENCES

- Martins PMM, Merfa MV, Takita MA, De Souza AA. Persistence in phytopathogenic bacteria: do we know enough? *Front Microbiol* 2018;9:1099. doi: 10.3389/fmicb.2018.01099
- Yang LN, He MH, Ouyang HB, Zhu W, Pan ZC, Sui QJ, Shang LP, Zhan J. Cross-resistance of the pathogenic fungus *Alternaria alternata* to fungicides with different modes of action. *BMC Microbiol* 2019;19:205. doi: 10.1186/s12866-019-1574-8
- Ramezani Y, Taheri P, Mamarabadi M. Identification of *Alternaria* spp. associated with tomato early blight in Iran and investigating some of their virulence factors. *J Plant Pathol* 2019;101:647–59. doi: 10.1007/s42161-019-00259-w
- Chaerani R, Voorrips RE. Tomato early blight (*Alternaria solani*): the pathogen, genetics, and breeding for resistance. *J Gen Plant Pathol* 2006;72:335–47. doi: 10.1007/s10327-006-0299-3
- Escrivá L, Oueslati S, Font G, Manyes L. *Alternaria* mycotoxins in food and feed: An overview. *J Food Quality* 2017;2017:1569748. doi: 10.1155/2017/1569748
- Shugar MA, Montgomery WW, Hyslop NE Jr. *Alternaria* sinusitis. *Ann Otol Rhinol Laryngol* 1981;90:251–4. doi: 10.1177/000348948109000311

7. Zahra LV, Mallia D, Hardie JG, Bezzina A, Fenech T. Case report. Keratomycosis due to *Alternaria alternata* in a diabetic patient. *Mycoses* 2002;45:512–4. doi: 10.1046/j.1439-0507.2002.00806.x
8. Chhabra V, Rastogi S, Barua M, Kumar S. *Alternaria alternata* infection associated osteomyelitis of maxilla: a rare disease entity. *Indian J Dent Res* 2013;24:639–41. doi: 10.4103/0970-9290.123420
9. Hattab Z, Ben Lasfar N, Abid M, Bellazreg F, Fathallah A, Hachfi W, Letaief A. *Alternaria alternata* infection causing rhinosinusitis and orbital involvement in an immunocompetent patient. *New Microbes New Infect* 2019;32:100561. doi: 10.1016/j.nmni.2019.100561
10. European Food Safety Authority (EFSA). Scientific Opinion on the risks for animal and public health related to the presence of *Alternaria* toxins in feed and food. EFSA Panel on Contaminants in the Food Chain (CONTAM). *EFSA J* 2011;9(10):2407. doi: 10.2903/j.efsa.2011.2407
11. Avenot HF, Michailides TJ. Resistance to boscalid fungicide in *Alternaria alternata* isolates from pistachio in California. *Plant Dis* 2007;91:1345–50. doi: 10.1094/pdis-91-10-1345
12. Ma Z, Felts D, Michailides TJ. Resistance to azoxystrobin in *Alternaria* isolates from pistachio in California. *Pestic Biochem Physiol* 2003;77:66–74. doi: 10.1016/j.pestbp.2003.08.002
13. Viriyasuthee W, Jogloy S, Saksirirat W, Saepaisan S, Gleason ML, Chen RS. Biological control of *Alternaria* leaf spot caused by *Alternaria* spp. in Jerusalem artichoke (*Helianthus tuberosus* L.) under two fertilization regimes. *Plants* 2019;8(11):463. doi: 10.3390/plants8110463
14. Adamczyk-Woźniak A, Komarovska-Porokhnyavets O, Misterkiewicz B, Novikov VP, Sporzyński A. Biological activity of selected boronic acids and their derivatives. *Appl Organometal Chem* 2012;26:390–3. doi: 10.1002/aoc.2880
15. Hall DG. Structure, properties, and preparation of boronic acid derivatives. In: Hall DG, editor. *Boronic acids: preparation and applications in organic synthesis, medicine and materials*, 1&2. 2nd ed. Weinheim: Wiley-VCH; 2011. doi: org/10.1002/9783527639328.ch1
16. Martinko K, Tolvajčić P, Đermić E, Đermić D. Determination of minimum inhibitory concentration of boric acid on phytopathogenic bacterium *Pseudomonas tomato*. In: Rozman V, Antunović Z, editors. *Proceedings of 56th Croatian and 16th International Symposium on Agriculture*; 5–10 September 2021; Vodic, Croatia. Osijek: VIN Grafika; 2021. p. 393–7.
17. Michaelis A, Becker B. Ueber Monophenylborchlorid und die Valenz des Bors [About monophenylboron chloride and the valence of boron, in German]. *Ber Dtsch Chem Ges* 1880;13:58–61. doi: 10.1002/cber.18800130118
18. Michaelis A, Becker B. Ueber Monophenylborchlorid und einige Derivate desselben [About monophenylboron chloride and some of its derivatives, in German]. *Ber Dtsch Chem Ges* 1882;15:180–5. doi: 10.1002/cber.18820150143
19. Marasović M, Ivanković S, Stojković R, Đermić D, Galić B, Miloš M. *In vitro* and *in vivo* antitumour effects of phenylboronic acid against mouse mammary adenocarcinoma 4T1 and squamous carcinoma SCCVII cells. *J Enzyme Inhib Med Chem* 2017;32:1299–304. doi: 10.1080/14756366.2017.1384823
20. Obuobi S, Voo ZX, Low MW, Czarny B, Selvarajan V, Ibrahim NI, Yang YY, Ee PLR. Phenylboronic acid functionalized polycarbonate hydrogels for controlled release of polymyxin B in *Pseudomonas aeruginosa* infected burn wounds. *Adv Healthc Mater* 2018;7(13):1701388. doi: 10.1002/adhm.201701388
21. Ferrer-Espada R, Sánchez-Gómez S, Pitts B, Stewart PS, Martínez-de-Tejada G. Permeability enhancers sensitize β -lactamase-expressing *Enterobacteriaceae* and *Pseudomonas aeruginosa* to β -lactamase inhibitors thereby restoring their β -lactam susceptibility. *Int J Antimicrob Agents* 2020;56:105986. doi: 10.1016/j.ijantimicag.2020.105986
22. Yalinkiliç MK, Yoshimura T, Takahashi TYM. Enhancement of the biological resistance of wood by phenylboronic acid treatment. *J Wood Sci* 1998;44:152–7. doi: 10.1007/BF00526262
23. Liu X, Laks PE, Pruner MS. A preliminary report on the wood preservative properties of phenylboronic acid. *Forest Prod J* 1994;44:46–8.
24. Molodykh ZV, Teplyakova LV, Nikonov GN, Erastov O. Fungitsidnaya aktivnost' proizvodnykh difenilbornoj kisloty [Fungicidal activity of diphenylboric acid derivatives, in Russian]. *Fiziol Akt Veshchestva* 1988;20:68–71.
25. Freeman A, Segal R, Dror Y. Methods and compositions for treating fungal infections. US patent. United States Patent No. 7,825,104 B2, 2010 [displayed 15 March 2022]. Available at <https://patentimages.storage.googleapis.com/4d/44/bc/505ed941ccea41/US7825104.pdf>
26. U.S. Environmental Protection Agency. Boronic acid, phenyl-98-80-6 | DTXSID9059179 Searched by DTXSID9059179 [displayed 13 March 2022]. Available at <https://comptox.epa.gov/dashboard/chemical/details/>
27. Soriano-Ursúa MA, Farfán-García ED, López-Cabrera Y, Querejeta E, Trujillo-Ferrara JG. Boron-containing acids: preliminary evaluation of acute toxicity and access to the brain determined by Raman scattering spectroscopy. *Neurotoxicology* 2014;40:8–15. doi: 10.1016/j.neuro.2013.10.005
28. Martinko K, Ivanković S, Lazarević B, Đermić E, Đermić D. Control of early blight fungus (*Alternaria alternata*) in tomato by boric and phenylboronic acid. *Antibiotics* 2022;11(3):320. doi: 10.3390/antibiotics11030320
29. Brown PH, Bellaloui N, Wimmer MA, Bassil ES, Ruiz J, Hu H, Pfeffer H, Dannel F, Römheld V. Boron in plant biology. *Plant Biol* 2002;4:205–23. doi: 10.1055/s-2002-25740
30. Nagrale DT, Gaikwad AP, Sharma L. Morphological and cultural characterization of *Alternaria alternata* (Fr.) Keissler blight of gerbera (*Gerbera jamesonii* H. Bolus ex J.D. Hook). *J Appl Nat Sci* 2013;5:171–8. doi: 10.31018/jans.v5i1.302
31. Zheng HH, Zhao J, Wang TY, Wu XH. Characterization of *Alternaria* species associated with potato foliar diseases in China. *Plant Pathol* 2015;64:425–33. doi: 10.1111/ppa.12274
32. Grover RK, Moore JD. Toximetric studies of fungicides against brown rot organism *Sclerotinia fruticola*. *Phytopathology* 1962;52:876–9.
33. Qadoos M, Kahn MI, Suleman M, Khan H, Aqeel M, Rafiq M. Comparison of poison food technique and drench method for *in vitro* control of *Alternaria* sp., the cause of leaf spot of bitter melon. *Merit Res J Agric Sci Soil Sci* 2016;4:126–30.
34. Schneider CA, Rasband WS, Eliceiri KW. NIH Image to ImageJ: 25 Years of image analysis. *Nat Methods* 2012;9:671–5. doi: 10.1038/nmeth.2089
35. Guzmán C, Bagga M, Kaur A, Westermarck J, Abankwa D. ColonyArea: an ImageJ plugin to automatically quantify colony formation in clonogenic assays. *PLoS One* 2014;9(3):e92444. doi: 10.1371/journal.pone.0092444
36. Singh RS, Khanna RN. Effect of certain inorganic chemicals on growth and spore germination of *Alternaria tenuis* auct., the fungus causing

- core rot of mandarin oranges in India. Mycopathol Mycol Appl 1969;37:89–96. doi: 10.1007/BF02051337
37. Patel NA, Dange SRS, Patel S. Efficacy of chemicals in controlling fruit rot of tomato caused by *Alternaria tomato*. Indian J Agric Res 2005;39:72–5.
38. Bhalerao JB, Chavan RA, Dharbale BB, Kendre AH, Mete VS. Study on *in-vitro* efficacy of botanicals and chemicals against *Alternaria solani* associated with post-harvest rot of tomato (*Lycopersicon esculentum* Mill.). J Pharmacogn Phytochem 2019;8:2045–9.

In vitro* antimikotički učinak fenilboronske i borne kiseline na patogenu gljivu *Alternaria alternata

Askomicetna gljiva *Alternaria alternata* uzročnik je koncentrične pjegavosti, jedne od ekonomski najvažnijih bolesti rajčice. Zbog česte primjene fungicida, ta je gljiva razvila otpornost na agrokemikalije koje se koriste u njezinu suzbijanju, s negativnim ekonomskim i ekološkim posljedicama. Novi načini suzbijanja gljivičnih patogena uključuju upotrebu ekološki prihvatljivih i manje toksičnih spojeva, među koje potencijalno spadaju boronske kiseline. Pokusom *in vitro* istražen je antimikotički učinak fenilboronske i borne kiseline na gljivu *A. alternata*. Nakon izolacije patogena iz rajčice, određena je minimalna inhibitorna koncentracija fenilboronske i borne kiseline za rast micelija primjenom tehnike *poisoned food*. Antimikotički učinak testiran je na širokom rasponu koncentracija fenilboronske i borne kiseline (od 0,04 % do 0,3 %), pojedinačno umiješanih u hranjivu podlogu na kojima je tijekom petodnevnog inkubacije uzgajan micelarni disk kulture patogena. Fenilboronska je kiselina pri niskoj koncentraciji (0,05 %) potpuno inhibirala rast micelija. Primjena borne kiseline u različitim rasponima koncentracija nije značajno umanjila rast micelija, ali je primijećeno smanjenje sporulacije patogena, čime se potvrđuje fungistatski učinak borne kiseline. Prema našoj spoznaji, ovo je prva studija koja opisuje *in vitro* antimikotički učinak fenilboronske kiseline na patogen koji je važan u poljoprivredi. Štoviše, s obzirom na to da je *A. alternata* i patogen ljudi, studija ima i potencijalni medicinski značaj.

KLJUČNE RIJEČI: boronske kiseline; koncentrična pjegavost; minimalna inhibitorna koncentracija rajčica; sporulacija