Original article

Absence of mutations in the human interferon alpha-2b gene in workers chronically exposed to ionising radiation

Dauren Botbayev^{1,2,3}, Gloria Ravegnini², Giulia Sammarini², Polat Kazymbet⁴, Elisabetta Cilli⁵, Patrizia Serventi⁵, Alexandra Khanseitova¹, Bakhytzhan Alzhanuly¹, Ayaz Belkozhaev¹, Nagima Aitkhozhina¹, Meirat Bakhtin⁴, Vittorio Lodi⁶, Patrizia Hrelia³, and Sabrina Angelini³

¹ Aitkhozhin Institute of Molecular Biology and Biochemistry KS MES, Almaty, Kazakhstan

² Al-Farabi Kazakh National University, Almaty, Kazakhstan

³ Department of Pharmacy and Biotechnology, University of Bologna, Bologna, Italy

⁴ Institute of Radiobiology and Radiation Protection, Medical University, Astana, Kazakhstan

⁵ Laboratories of Physical Anthropology and Ancient DNA, Department of Cultural Heritage, University of Bologna, Ravenna Campus, Italy

⁶ Occupational Health Unit, Sant'Orsola-Malpighi Hospital, Bologna, Italy

[Received in September 2018; Similarity Check in September 2018; Accepted in March 2019]

Individuals chronically exposed to low-level ionising radiation (IR) run the risk of harmful and long-term adverse health effects, including gene mutations and cancer development. The search for reliable biomarkers of IR exposure in human population is still of great interest, as they may have a great implementation potential for the surveillance of occupationally exposed individuals. In this context, and considering previous literature, this study aimed to identify mutations in the human interferon alpha-2b (hIFN α -2b) as a potential biomarker of occupational chronic low-dose IR exposure linking low-IR exposure to the effects on haematopoiesis and reduced immunity. The analysis was performed in the genomic DNA of 51 uranium miners and 38 controls from Kazakhstan, and in 21 medical radiology workers and 21 controls from Italy. hIFN α -2b gene mutations were analysed with the real-time polymerase chain reaction (PCR) or Sanger sequencing. However, none of the investigated workers had the hIFN α -2b mutation. This finding highlights the need for further research to identify biomarkers for early detection of health effects associated with chronic low-dose IR exposure.

KEY WORDS: hIFNα-2b mutations; genotoxicity; radiology workers; uranium miners

In addition to cellular DNA damage and damage response, exposure to ionising radiation (IR) triggers nontargeted effects mainly related to the immune system (1-4). Two recent studies reported an association between mutations in the human interferon alpha-2b (hIFN α -2b) gene and chronic exposure to low-level of IR in medical personnel (5) and residents from Pakistan (6). The IFN α -2b gene encodes a protein belonging to the class of cytokines with immunomodulatory, antiviral, anti-proliferative, and anti-tumour activities (7–10). Mutations, typically the frameshift ones or single nucleotide changes in the human IFNα-2b gene could therefore compromise the functioning of the immune system. In addition, these mutations have been detected in brain tumour patients exposed to different environmental stressors, including high levels of naturally occurring IR (10), and other cancer patients (11, 12). We considered these findings interesting because of the health consequences associated with chronic exposure to low doses

of IR. As mutations in the human IFN α -2b gene might compromise immunity, we wondered if they could also serve as biomarkers for health monitoring of occupational and environmental chronic exposure to low IR doses. The aim of this study was therefore to verify the hypothesis in a group of uranium miners from Kazakhstan and in a group of Italian radiology workers.

MATERIALS AND METHODS

Study population

Volunteer coal miners, all men (n=51) were recruited from two Kazakh regions, Aksu and Zavodskoy (Northern Kazakhstan). In Aksu the mean effective dose is 4 mSv/ year, while in Zavodskoy it is 4.95 mSv/year (13). Figure 1 shows a map of the area with the sources of IR exposure. Matched controls (n=38) were anonymous blood donors recruited at the Almaty Blood Donation Centre. Formal written consent was signed by all participants before inclusion in the study.

Corresponding author: Sabrina Angelini, PhD, University of Bologna Department of Pharmacy and Biotechnology, Via Imerio 48, 40126 Bologna, Italy, E-mail: *s.angelini@unibo.it*



Figure 1 IR sources of exposure at locations of the Kazakh study arm

The Italian study arm consisted of 42 hospital workers, 21 of whom were occupationally exposed to IR and 21 were healthy, unexposed controls. These participants represent a subset of our previous study in radiological workers (14), whose DNA samples had been stored for future sub-studies. They had signed informed consent for the use of their samples in sub-studies such as this, according to Helsinki Declaration and its later amendments.

Both the Kazakh and Italian participants were taken 10 mL of blood in heparinised tubes during regular medical examination. Blood samples were stored at -80 °C until DNA isolation and subsequent IFN α -2b mutation analysis.

DNA isolation and IFNa-2b mutation analysis

Genomic DNA was isolated from frozen peripheral blood lymphocytes using a standardised phenol-chloroform extraction method. The DNA was purified with 70 % ethanol, air dried overnight at room temperature, and resuspended in approximately 100 μ L of deionised nuclease-free water. Before we analysed the samples for mutations, we selected ten IFN α -2b mutations appearing at the frequency higher than 5 %, as reported by Shahid et al. (5,

6), which, according to these authors, were associated with chronic low-dose IR exposure and its potential to induce point mutations in single genes and modulate the expression of a variety of genes (15–17).

We then analysed our samples for these ten mutations using real-time polymerase chain reaction (PCR) with a customised Applied Biosystems[™] Taqman Assay[®] (Thermo Fisher Scientific, Waltham, MA, USA) as recommended by manufacturer or using Sanger sequencing with an ABI Prism[™] 310 Genetic Analyzer (Thermo Fisher Scientific) as described by Shashid et al. (5) (Table 1). Blanks were included in each reaction for quality control.

RESULTS AND DISCUSSION

Surprisingly, none of the analysed samples had any of the ten mutations of this gene. Reasons for our negative findings may be several. One of them is a possible bias in the selection of mutations. Instead of sequencing the entire coding region of the hIFN α -2b gene, we selected ten most frequent mutations identified by Shahid et al. (5, 6). Another

Mutation (base position) [§]	Frequency*	Methods
C insertion (8–9)	5.4 %	Real-time PCR
A to T (187)	8 %	Real-time PCR
T to A (219)	8 %	Real-time PCR
A to G (256)	8 %	Real-time PCR
C insertion (330–331)	8 %	Real-time PCR
A deletion (435)	3.3 %	Sanger sequencing
G to A (436)	4.3 %	Sanger sequencing
A to G (437)	3.3 %	Sanger sequencing
A deletion (439)	4.3 %	Sanger sequencing
A deletion (477)	3.3 %	Real-time PCR

Table 1 Characteristics of the IFNa-2b gene mutation

* Frequency reported by Shahid et al (5, 6); § gene accession number NM_00605

reason could be the sample size, which is particularly small for the Italian arm. Yet another could be differences in genetic make-up, environmental background, lifestyle, and exposure between our study participants and the populations described by Shahid et al. (5, 6).

With regard to the Kazakh study arm, we did not have information about the effective annual dose received by each occupationally exposed worker participating in the study but only the mean doses for Aksu and Zavodskoy (4 and 4.95 mSv/year, respectively). While not knowing the exact magnitude of occupational exposure in this particular group may seriously limit the interpretation of our findings, the reported annual mean doses did not differ significantly from those reported by Shahid et al. (6).

As for the Italian arm, occupational exposure was routinely monitored by personal devices (film badges). The dose equivalent of IR to the whole body obtained from the personal dosimetry records ranged from 0.90 to 116.74 mSv (mean \pm standard deviation = 40.61 \pm 37.70 mSv) over the entire working life (which ranged from four to 34 years). Considering the wide range of the years of employment, the idea was also to investigate whether IFNa-2b mutations could identify a subgroup of occupationally exposed individuals more prone to IR-induced DNA damages as an important step towards a timely establishment of effective health surveillance programmes. However, none of the radiological workers showed mutation in the IFN α -2b gene. In addition, a routine occupational health examination showed no clinical or haematological abnormalities in any of the study participants (14). This differs from the report by Shahid et al. (5), in which complete blood count (CBC) revealed that radiation-exposed workers had more abnormal CBC findings than controls. Therefore, the lack of mutation in the Italian study group could also be due to a better health status of these workers. We also cannot exclude the involvement of a different ethnic susceptibility (i.e. different genetic make-up). DNA repair mechanisms may have an important role in eliminating the genetic stress caused by exposure to genotoxic agents, including IR. In this context, the presence of polymorphisms in DNA repair genes, which characterises individual genetic make-up, may contribute to different levels of mutational load and/or genetic damage as highlighted in several studies (18–24). With regard to genetic damage, our earlier studies (14, 19, 20), however limited by the small sample size, found significantly higher micronucleus (MN) frequency in radiological workers chronically exposed to low level of IR than in controls. These and other studies (19, 20, 22, 24) clearly confirm the association between occupational exposure to low-dose IR exposure and genotoxicity. MN formation is considered a reliable biomarker of exposure to clastogenic and aneugenic agents, including IR (26-28). In addition to MN, other cytogenetic tests, including the comet assay (29-32) and chromosome aberration assays (33–36), have confirmed the association between genotoxicity and low-dose radiation exposure.

CONCLUSIONS

Despite their promising potential, none of the available cytogenetic tests has become part of routine biodosimetry surveillance of occupationally and/or environmentally exposed individuals. These tests are traditionally manual and labour-intensive, even with the recently proposed automation protocol for MN and chromosomal aberration analysis (37, 38). Despite our enthusiasm about the promising potential of the IFN α -2b gene as a reliable biomarker of IR-associated immunological risk (5, 6), we did not observe any IFN α -2b gene mutation or changes in blood cell counts.

Unfortunately, we do not have any cytogenetic data (i.e. chromosome aberrations, MN, or the comet assay findings) or data about health consequences associated with low-dose radiation for the Kazakh study arm. In conclusion, our failure to find mutation in the IFN α -2b gene in either arm calls for further research that would identify reliable biomarkers for early detection of health effects associated with low-dose IR.

REFERENCES

- Godekmerdan A, Ozden M, Ayar A, Gursu FM, Ozan AT, Serhatlioglu S. Diminished cellular and humoral immunity in workers occupationally exposed to low levels of ionizing radiation. Arch Med Res 2004;35:324–8. doi: 10.1016/j. arcmed.2004.04.005
- Oskouii MR, Refahi S, Pourissa M, Tabarraei Y. Assessment of humoral immunity in workers occupationally exposed to low levels of ionizing radiation. Life Sci J 2013;10:58–62.
- Rödel F, Frey B, Multhoff G, Gaipl U. Contribution of the immune system to bystander and non-targeted effects of ionizing radiation. Cancer Lett 2015;356:105–13. doi: 10.1016/j.canlet.2013.09.015
- Voos P, Fuck S, Weipert F, Babel L, Tandl D, Meckel T, Hehlgans S, Fournier C, Moroni A, Rödel F, Thiel G. Ionizing radiation induces morphological changes and immunological modulation of Jurkat cells. Front Immunol 2018;9:922. doi: 10.3389/fimmu.2018.00922
- Shahid S, Mahmood N, Nawaz Chaundhry M, Sheikh S, Ahmad N. Mutations of the human interferon alpha-2b (hIFNα2b) gene in occupationally protracted low dose radiation exposed personnel. Cytokine 2015;73:181–9. doi: 10.1016/j. cyto.2015.02.008
- Shahid S, Mahmood N, Nawaz Chaundhry M, Ahmad N. Mutations of the human interferon alpha-2b (hIFN-α2b) gene in low-dose natural terrestrial ionizing radiation exposed dwellers. Cytokine 2015;76:294–302. doi: 10.1016/j. cyto.2015.05.011
- Parmar S, Platanis LC. Interferons: mechanisms of action and clinical implications. Curr Opin Oncol 2003;15:431–9. PMID: 14624225
- Brandacher G, Winkler C, Schroecksnadel K, Margreiter R, Fuchs D. Antitumoral activity of interferon-gamma involved in impaired function in cancer patients. Curr Drug Metab 2006;7:599–612. doi: 10.2174/138920006778017768

- Bose A, Baral R. IFNα2b stimulated release of IFNgamma differentially regulates T cell and NK cell mediated tumor cell cytotoxicity. Immunol Lett 2007;108:68–77. doi: 10.1016/j. imlet.2006.10.002
- Levin D, Schneider WM, Hoffmann HH, Yarden G, Busetto AG, Manor O, Sharma N, Rice CM, Schreiber G. Multifaceted activities of type I interferon are revealed by a receptor antagonist. Sci Signal 2014;7:ra50. doi: 10.1126/ scisignal.2004998
- Shahid S, Nawaz Chaudry M, Mahmood N, Sheikh S. Mutation of the human interferon alpha-2b gene in brain tumor patients exposed to different environmental conditions. Cancer Gene Ther 2015;22:246–61. doi: 10.1038/cgt.2015.12
- Shahid S, Nawaz Chaundhry M, Mahmood N. Mutations of the human interferon alpha-2b (hIFNα-2b) gene in cancer patients receiving radiotherapy. Am J Cancer Res 2015;5:2455– 66. PMCID: PMC4568781
- Kazymbet PK, Bakhtin MM, Imasheva BS. Population radiation level of the north Kazakhstan by natural sources of ionized radiation. Astana Med J 2006;1:26-8.
- Maffei F, Angelini S, Forti GC, Lodi V, Mattioli S, Hrelia P. Micronuclei frequencies in hospital workers occupationally exposed to low levels of ionizing radiation: influence of smoking status and other factors. Mutagenesis 2002;17:405– 9. doi: 10.1093/mutage/17.5.405
- Trott K, Rosemann M. Molecular mechanisms of radiation carcinogenesis and the linear, non-threshold dose response model of radiation risk estimation. Radiat Environ Biophys 2000;39:79–87. PMID: 10929376
- Gadhia P, Shah N, Nahata S, Patel S, Patel K, Pithawala M, Tamakuwala D. Cytogenetic analysis of radiotherapeutic and diagnostic workers occupationally exposed to radiations. Int J Human genet 2004;4:65. doi. 10.1080/09723757. 2004.11885872
- 17. Jin YW, Na YJ, Lee YJ, An S, Lee JE, Jung M, Kim H, Nam SY, Kim CS, Yang KH, Kim SU, Kim WK, Park WY, Yoo KY, Kim CS, Kim JH. Comprehensive analysis of time-and dose-dependent patterns of gene expression in a human mesenchymal stem cell line exposed to low-doses ionizing radiation. Oncol Rep 2008;19:135–44. doi: 10.3892/or.19.1.135
- Kazakh Ministry of Justice, Centre of Legal Information. О Стратегическом плане Агентства Республики Казахстан по атомной энергии на 2012 – 2016 годы [Strategic Plan of the Atomic Energy Agency of the Republic of Kazakhstan for 2012–2016, in Russian]. [displayed 27 March 2019]. Available at http://www.adilet.zan.kz/rus/docs/P1200001806/links
- Angelini S, Kumar R, Carbone F, Maffei F, Cantelli-Forti G, Violante FS, Lodi V, Curti S, Hemmini K, Hrelia P. Micronuclei in humans induced by exposure to low level of ionizing radiation: influence of polymorphismsin DNA repair genes. Mutat Res 2005;570:105–17. doi: 10.1016/j. mrfmmm.2004.10.007
- Milić M, Rozgaj R, Kašuba V, Jazbec AM, Starčević B, Lyzbicki B, Ravegnini G, Zenesini C, Musti M, Hrelia P, Angelini S. Polymorpisms in DNA repair genes: link with biomarkers of the CBMN cytome assay in hospital workers chronically exposed to low doses of ionising radiation. Arh Hig Rada Toksikol 2015;66:109–20. doi: 10.1515/aiht-2015-66-2655
- Mumbrekar KD, Goutham HV, Vadhiraja BM, Bola Sadashiva SR. Polymorphisms in double strand break repair related genes influence radiosensitivity phenotype in lymphocytes from

healthy individuals. DNA Repair 2016;40:27-34. doi: 10.1016/j.dnarep.2016.02.006

- Sinitsky MY, Minina VI, Asanov MA, Yuzhalin AE, Ponasenko AV, Druzhinin VG. Association of DNA repair gene polymorphisms with genotoxic stress in underground coal miners. Mutagenesis 2017;32:501–9. doi: 10.1093/mutage/ gex018
- Doukali H, Ben Salah G, Ben Rhouma B, Hajjaji M, Jaouadi A, Belguith-Mahfouth N, Masmoudi ML, Ammar-Keskes L, Kamoun H. Cytogenetic monitoring of hospital staff exposed to ionizing radiation: optimize protocol considering DNA repair genes variability. Int J Radiat Biol 2017;93:1283–8. doi: 10.1080/09553002.2017.1377361
- Angelini S, Kumar R, Carbone F, Bermejo JL, Maffei F, Cantelli-Forti G, Hemminki K, Hrelia P. Inherited susceptibility to bleomycin-induced micronuclei: Correlating polymorphisms in GSTT1, GSTM1 and DNA repair genes with mutagen sensitivity. Mutat Res 2008;638:90–7. doi: 10.1016/j. mrfmmm.2007.09.001
- 25. Milić M, Rozgaj R, Kašuba V, Jazbec AM, Hrelia P, Angelini S. The influence of individual genome sensitivity in DNA damage repair assessment in chronic professional exposure to low doses of ionizing radiation. In: Chen CC, editor. Selected topics in DNA repair. London: IntechOpen; 2011 [displayed 27 March 2019]. Available at https://www.intechopen.com/ books/selected-topics-in-dna-repair/the-influence-of-individual-genome-sensitivity-in-dna-damage-repair-assessment-in-chronic-profession
- 26. Fenech M, Knasmueller S, Bolognesi C, Bonassi S, Holland N, Migliore L, Palitti F, Natarajan AT, Kirsch-Volders M. Molecular mechanisms by which *in vivo* exposure to exogenous chemical genotoxic agents can lead to micronucleus formation in lymphocytes *in vivo* and *ex vivo* in humans. Mutat Res 2016;770:12–25. doi: 10.1016/j.mrrev.2016.04.008
- 27. Angelini S, Bermejo JL, Ravegnini G, Sammarini G, Hrelia P. Application of the lymphocyte Cytochinesis-Block Micronucleus Assay to population exposed to petroleum and its derivatives: results from a systematic review and metaanalysis. Mutat Res 2016;770:58–72. doi: 10.1016/j. mrrev.2016.03.001
- Siama Z, Zosang-Zuali M, Vanlalruati A, Jagetia GC, Pau KS, Kumar NS. Chronic low dose exposure of hospital workers to ionizing radiation leads to incressed micronuclei frequency and reduced antioxidants in their peripheral blood lymphocytes. Int J Radiat Biol 2019. doi: 10.1080/09553002.2019.1571255. [Epub ahead of print]
- Khisroon M, Khan A, Naseem M, Ali N, Khan S, Rasheed SB. Evaluation of DNA damage in lymphocytes of radiology personnel by comet assay. J Occup Health 2015;57:268–74. doi: 10.1539/joh.14-0154-OA
- 30. Korzeneva IB, Kostuvk SV, Ershova LS, Osipov AN, Zhuraleva VF, Pankratova GV, Porokhovnik LN, Veiko NN. Human circulating plasma DNA significantly decreases while lymphocyte DNA damage increases under chronic occupational exposure to low-dose gamma-neutron and tritium β -radiation. Mutat Res 2015;779:1–15. doi: 10.1016/j.mrfmmm.2015.05.004
- Gulati S, Yadav A, Kumar N, Kanupriya, Aggarwal NK, Kumar R, Gupta R. Effect of GSTM1 and GSTT1 polymorphisms on genetic damage in humans population exposed to radiation from mobile towers. Arch Environ Contam Toxicol 2016;70:615–25. doi: 10.1007/s00244-015-0195-y

- Gaetani S, Monaco F, Bracci M, Ciarapica V, Impollonia G, Valentino M, Tomasetti M, Santarelli L, Amati M. DNA damage response in workers exposed to low-dose ionising radiation. Occup Environ Med 2018;75:724–9. doi: 10.1136/ oemed-2018-105094
- Maffei F, Angelini S, Forti GC, Violante FS, Lodi V, Mattioli S, Hrelia P. Spectrum of chromosomal aberrations in peripheral lymphocyte of hospital workers occupationally exposed to low doses of ionizing radiation. Mutat Res 2004;547:91–9. doi: 10.1016/j.mrfmmm.2003.12.003
- 34. Tawn EJ, Curwen GB, Jonas P, Gillies M, Hodgson L, Cadwell KK. Chromosome aberrations determined by FISH in radiation workers from the Sellafield nuclear facility. Radiat Res 2015;184:296–303. doi: 10.1667/RR14125.1
- 35. Djokovic-Davidovic J, Milovanovic A, Milovanovic J, Antic V, Gajic M. Analysis of chromosomal aberrations frequency,

haematological parameters and received doses by nuclear medicine professionals. J BUON 2016;21:1307–15. PMID: 27837637

- Tawn EJ, Curwen GB, Riddel AE. Chromosome aberrations in workers occupationally exposed to tritium. J Radiol Prot 2018;38:N9–16. doi: 10.1088/1361-6498/aab0d0
- Shirley B, Li Y, Knoll JHM, Rogan PK. Expedite radiation biodosimetry by automated dicentric chromosome identification (ADCI) and dose estimation. J Vis Exp 2017;127:56245. doi: 10.3791/56245
- Lenzi M, Cocchi V, Hrelia P. Flow cytometry vs optical microscopy in the evaluation of the genotoxic potential of xenobiotic compounds. Cytometry B Clin Cytom 2018;94:696– 706. doi: 10.1002/cyto.b.21546

Izostanak mutacija humanoga interferon alfa-2b gena u radnika kronično izloženih ionizirajućem zračenju

Kronična izloženost niskim razinama ionizirajućega zračenja povezana je s rizikom od dugoročnih štetnih posljedica za zdravlje, što obuhvaća i mutacije gena te nastanak raka. U tijeku je potraga za pouzdanim biopokazateljima izloženosti ionizirajućem zračenju u ljudi, budući da njihova primjena može značajno unaprijediti praćenje profesionalno izloženih osoba. U tom smislu, a s obzirom na ranija saznanja, cilj je ovoga istraživanja bio utvrditi mutacije gena za proizvodnju humanoga interferona alfa-2b (hIFN α -2b gena) kao mogućega biopokazatelja profesionalne kronične izloženosti niskim dozama ionizirajućega zračenja, koje je usto povezano s djelovanjem na hematopoezu i pad imuniteta. Analiziran je genomski DNA 51 rudara u rudnicima uranija te 38 kontrolnih ispitanika iz Kazahstana, odnosno genomski DNA 21 zdravstvenoga radnika na radiologiji i 21 kontrolnoga ispitanika iz Italije. Mutacije hIFN α -2b gena utvrđivane su metodom lančane reakcije polimerazom u stvarnom vremenu (engl. *real-time PCR*) odnosno sekvenciranjem prema Sangeru, ali se pokazalo da niti jedan radnik nije imao niti jednu od deset traženih mutacija toga gena. Stoga ne preostaje drugo nego i dalje tražiti pouzdane biopokazatelje za rano otkrivanje štetnih zdravstvenih učinaka povezanih s kroničnom izloženosti niskim dozama ionizirajućega zračenja.

KLJUČNE RIJEČI: hIFNα-2b; genotoksičnost; radiologija; rudari, uranij; zdravstveni radnici